Anticancer Activity of Metal Complexes with Acesulfame Mixed with Triphenylphosphine Ligands

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ABSTRACT

Metal complexes of silver(I), nickel(II) and copper(II) with the mixed ligands of acesulfame (ace) and triphenylphosphine (PPh₂) were prepared in this work. The complexes of $[Ag(ace)]_n$ (1), $[Ag(ace)(PPh_3)]$ (2), $[Ni(ace)_{(H_2O)_4}]$ (3), $[Ni(ace)_{(PPh_2)_2}]$ (4), $[Cu(ace)_{(H_2O)_2}]$ (5) and [Cu(ace),(PPh,),] (6) were characterised by FT-IR and NMR spectroscopy, Mass spectrometry including X-Ray powder diffraction. The synthesised complexes were examined in their anticancer activity against A549 lung cancer cells which were further evaluated their efficiency by IC_{50} method. $[Ag(ace)]_{u}$ (1), $[Ni(ace)_{u}(H,O)_{d}]$ (3) and $[Cu(ace)_{u}(H,O)_{d}]$ (5) which contain only acesulfame ligand exhibited higher IC_{50} values in comparison to the mixed ligands of ace and PPh, [Ag(ace)(PPh,)] (2), [Ni(ace),(PPh,),] (4) and [Cu(ace),(PPh,),] (6) displayed the inhibition in the growth of cancer cells with IC_{50} values of 1.65, 13.45 and 1.06 μ M, respectively. Additionally, silver(I) and copper(II) complexes also presented better activity than the standard Etoposide (IC₅₀ 5.623 μ M). The results indicated that the mixed ligands could perform more excellent inhibition of cancer cells growing in comparison to the using of only acesulfame or triphenylphosphine ligand.

Keywords: Complexes, Acesulfame, Anticancer, Triphenylphosphine

INTRODUCTION

Anticancer activity is one of the most attractive applications involved the examination in capability of coordination compounds in biological studies. Silver(I) complexes was found to be the most effective substrate in antibacterial and antifungal studies for several years (Kumar et al., 2010; Chandra and Ruchi, 2013; Charef et al., 2015. Our recent research also revealed that silver(I) complexes incorporated with the mixed ligands of triphenylphosphine and acesulfame exhibit excellent activity against A549 lung cancer cells (Boonseng et al., 2018). Herein, we would like to expand the scope of metals to the first-row transition metals; nickel(II) and copper(II) which has been prevalently used for numerous catalysis and biological studies (Chandra et al., 2015; Ghobadi et al., 2018; Mo et al., 2018; Saha et al., 2018).

Acesulfame-K is a commercially artificial sweetener in which its anionic form is considered to be an excellent ligand according to the containing of several donor atoms. Apart from conveniently and commercially available, the compound was also found to provide low toxicity at low concentration leading to the practically using for living organism (Cong et al., 2013). Owing to the prior advantage of acesulfame-K, the recent research by Cavicchioli and co-workers (2010) was revealed that the using of acesulfame as the ligand for platinum(II) and silver(I) complexes exhibited exceptional antimycobacterial and antibacterial activity. Moreover, in order to improve the ability of metal complexes in biological activities regarding the inhibition of cancer cells growing, several researches have been attracted to the structural modification of transition metal complexes. The study by the Nawaz group (2011) revealed that the introduction of triphenylphosphine (PPh,) into the heterocyclic thiones silver(I) complexes could perform more excellent activity against cancer cells than using only thione compounds. Thereafter, Kaplan and co-workers reported the activity of silver(I) complex against lung cancer A549 cells via MTT assay in which the better reactivity of such complex than its coordinated ligands were discovered (Kaplan, 2017). Lung cancer has become our desired target according to the report by World Health Organization (WHO) which indicated that lung cancer is the most common cause of cancer-related death of people and more than 50 percent of lung cancer cases occurred in less developed countries (Gazdar et al., 2010).

Therefore, in this study, the complexes of silver(I), nickel(II) and copper(II) incorporated with the mixed ligands of acesulfame and PPh₃ were synthesized and their structures were determined by FT-IR, ¹H and ³¹P NMR spectroscopy, mass spectrometry including X-Ray powder diffraction. The products were examined in their inhibition of lung cancer cells growing (A549) and their abilities were presented and compared in term of IC₅₀ values.

MATERIALS AND METHODS

Materials

All the chemicals used in this study were at least analytical grade. Silver nitrate (AgNO₃) was obtained from Labscan. NiCl₂(PPh₃)₂, Ni(CH₃CO₂)₂·4H₂O and Cu(CH₃CO₂)₂·xH₂O were purchased from Aldrich. Triphenylphosphine (PPh₃) and acesulfame-K (ace-K) were supplied by Fluka. Other solvents were used without further purification. The IR spectra were recorded in wavenumber region 4,000-400 cm⁻¹on a PerkinElmer FT-IR spectrophotometer. The ¹H and ³¹P NMR spectra in DMSO- d_6 were operated at the frequency of 400 MHz (Bruker). Mass spectrometric analysis was performed using the ESI technique (Agilent 6540 LC/MS system). X-Ray powder diffraction was performed using diffractometer provided by Rigaku (MiniflexII). [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] (MTT) assay was analysed by measurement the absorbance of formarzan product using Microplate Reader from PerkinElmer, Spectrum GX.

Synthesis of metal complexes with acesulfame and mixed ligands of acesulfame and triphenylphosphine

Silver(I) complexes. $[Ag(ace)]_n$ (1) was prepared according to the literature (Cavicchioli et al., 2010). 2.0 mmol of AgNO₃ was dissolved in water 25 mL and was then slowly added to the aqueous solution of acesulfame-K (2.0 mmol, 25 mL) under stirring. The mixture solution was left in the absence of light for slow evaporation for 2 days and the colorless crystals were obtained. $[Ag(ace)(PPh_3)]$ (2) was synthesized by the addition of AgNO₃ (1.0 mmol) aqueous solution into the ethanolic solution of PPh₃ (1.0 mmol). After stirring at 80 °C for 2 hours, acesulfame-K (1.0 mmol) in water was added into the mixture with continuously stirring at room temperature for 5 hours. The obtained product was filtered and recrystallized with ethanol.

Nickel(II) complexes. $[Ni(ace)_2(H_2O)_4]$ (3) was prepared according to the literature (Icbudak *et al.*, 2006) by adding drop-wise of an aqueous solution of acesulfame-K (2.0 mmol) 50 mL into a hot solution of Ni(CH₃CO₂)₂·4H₂O (1.0 mmol) in 50 ml ethanol with continuously stirring. The mixture was stirred at 80 °C for 2 hours. The green powder was achieved after the evaporation of solvent. $[Ni(ace)_2(PPh_3)_2]$ (4) was synthesized by adding the ethanolic solution of acesulfame-K (2.0 mmol) into the round bottom flask containing NiCl₂(PPh₃)₃ (1.0 mmol) in ethanol. The mixture was kept under reflux for 3 hours, then the reaction was cooled down to room temperature. The desired product was obtained by filtration and the filtrate was removed its solvent by rotary evaporator, the pale-green solid was yielded.

Copper(II) complexes. $[Cu(ace)_2(H_2O)_2]$ (5) was synthesized with the same procedure as complex $[Ni(ace)_2(H_2O)_4]$ (3) using $Cu(CH_3CO_2)_2 \cdot xH_2O$ instead of $Ni(CH_3CO_2)_2 \cdot 4H_2O$. The product was obtained as pale-blue solid. $[Cu(ace)_2(PPh_3)_2]$ (6) was prepared by mixing $Cu(CH_3CO_2)_2 \cdot xH_2O$ and two equivalents of acesulfame-K and PPh₃. The reaction was set up in the same condition as for complex $[Ag(ace)(PPh_3)]$ (2) to yield blue solid of the product.

Anticancer studies

The anticancer activity of the synthesized complexes was investigated against A549 lung cancer cells and further evaluated by IC₅₀ method. A549 cells (about 5,000 cells/well) were incubated in complete medium (10% fetal bovine serum +1% Penicillin Streptomycin in modified Eagle's medium) at 37 °C for 8 hours. The synthesized complexes were dissolved in water or dimethylsulfoxide with concentration of 50 mM and then a series of diluted solutions was added in triplicate into each well. The cells were incubated at 37 °C for 48 hours and after that the added media were aspirated. The MTT reagent (1% in growth media) 100 µL was added to each well and incubated at 37 °C for 4 hours. After the removal of MTT solution, 100 µL of dimethylsulfoxide was added to observe the absorbance at $\lambda_{max} = 570$ nm. The results were calculated for IC₅₀ values using GraphPad Prism7 software and compared with standard Etoposide.

RESULTS

Synthesis of metal complexes with acesulfame and mixed ligands of acesulfame and triphenylphosphine

The complexes of silver(I), nickel(II) and copper(II) were prepared as aforementioned in material and methods section. Melting points of complexes were determined at the heating rate of 5 °C/min using GALLENKAMP Melting Point Apparatus. Appearance and percentage yield of each compound are shown in Table 1.

Compounds	Appearance	%Yield	Melting point (°C)
$\left[\operatorname{Ag}(\operatorname{ace})\right]_{n}(1)$	White powder	46.3	202.3
$[Ag(ace)(PPh_3)] (2)$	White powder	62.7	190.3
$[Ni(ace)_2(H_2O)_4]$ (3)	Green powder	72.0	94.3
$[Ni(ace)_2(PPh_3)_2]$ (4)	Green powder	42.1	90.8
$[Cu(ace)_2(H_2O)_2]$ (5)	Pale-blue powder	50.8	262.2
$[Cu(ace)_2(PPh_3)_2](6)$	Blue powder	36.3	217.7

Table 1. General information of the synthesized complexes.

Characterization of metal complexes with acesulfame and mixed ligands of acesulfame and triphenylphosphine

The obtained complexes were analyzed in compositions and structures by IR and NMR spectroscopic methods, mass spectrometry including X-Ray powder diffraction to acquire molecular structures of the synthesized products.

IR studies

Acesulfame-K ligand displays frequencies at wavenumbers of 1,651, 1,588 and 1,287 cm⁻¹ which are corresponding to the vibration of C=O, S(=O)₂ and C-N functional groups, respectively. The frequency changes upon the complex formation of copper(II) ion are presented as the spectra shown in Figure 1. The wavenumbers at 3,565 cm⁻¹ and 3,431 cm⁻¹ of $[Cu(ace)_2(H_2O)_2]$ (5) are represented the stretching vibrations of O-H functional group of water molecules and these vibrations are also displayed in $[Ni(ace)_2(H_2O)_4]$ (3). Additionally, the carbonyl; C=O, and the sulfone; S(=O)₂ groups of complex 5 also exhibit frequencies at lower wavenumbers, 1,646 and 1,580 cm⁻¹, in comparison to the free acesulfame compound. Sharp bands are observed at wavenumbers of 527 and 516 cm⁻¹ in complex 5 and wavenumbers of 518 and 644 cm⁻¹ in complex 5 shows the donation of water molecule towards the metal center whilst the assigned wavenumber of complex 6 belongs to M-P bond.



Figure 1. IR spectra of $[Cu(ace)_2(H_2O)_2]$ (5) and $[Cu(ace)_2(PPh_3)_2]$ (6).

In order to compare the different types of metal centers affected the vibration frequencies, complexes 2, 4 and 6 are chosen and their spectra are presented in Figure 2. A high frequency shift in v(C-N) bands in all complexes indicate the coordination of ligand to the metal ions. M-N bond was observed in the region of 559-516 cm⁻¹ whilst M-O bond was also observed according to the metal-oxygen of water or metal-oxygen of acesulfame molecule. Moreover, the frequencies involved M-P bond formation in complexes 2, 4 and 6 were exhibited at the wavenumbers about 660-640 cm⁻¹. In addition, an unexpected signal was observed in complex 4 at the frequencies range of 3,400-3,050 cm⁻¹ which might be attributed to the moisture contained in the substrate. IR data of free acesulfame ligand and all complexes are shown in Table 2.



Figure 2. IR spectra of $[Ag(ace)(PPh_3)]$ (2), $[Ni(ace)_2(PPh_3)_2]$ (4) and $[Cu(ace)_2(PPh_3)_2]$ (6).

Compounds	v(OH)	v(C=O)	$v(S(=O)_2)$	v(C-N)	v(M-N)	v(M-O)/(M/P)
Acesulfame	-	1,651	1,588	1,287	-	-
$\left[Ag(ace)\right]_{n}$ (1)	-	1,644	1,563	1,312	559	513 (M-O)
$[Ag(ace)(PPh_3)] (2)$	-	1,643	1,566	1,342	557	644 (M-P)
[Ni(ace) ₂ (H ₂ O) ₄](3)	3,465, 3,364	1,638	1,557	1,323	530	517 (M-O)
$[Ni(ace)_2(PPh_3)_2](4)$	3,400- 3,050	1,644	1,548	1,334	538	645 (M-P)
$[Cu(ace)_2(H_2O)_2](5)$	3,565, 3,431	1,646	1,580	1,291	527	516 (M-O)
$[Cu(ace)_2(PPh_3)_2](6)$	-	1,647	1,549	1,329	518	664 (M-P)

Table 2. IR data of free ligands and complexes 1-6.

NMR studies

The synthesized complexes were observed in ¹H and ³¹P NMR spectra to examine the change of proton and phosphorus signals upon the coordination of ligands to the metal ions.

Generally, free PPh₃ shows the ³¹P signal at the chemical shift about -5.25 ppm. For the synthesized complexes **2**, **4** and **6**, the observed phosphorus atom of each one was found to resonate at the chemical shift of 15.81, 25.62 and 25.50 ppm, respectively. Complex **2** has been already known in its molecular structure from X-ray single crystal diffraction previously reported by our group. Structure of [Ag(ace)(PPh₃)] (**2**) is shown in Figure 3.



Figure 3. The molecular structure of [Ag(ace)(PPh₃)] (2).

However, complexes **4** and **6** still cannot be affirmed the exact position of PPh₃ coordinated to nickel(II) and copper(II) ions. To the extent of our knowledge including the steric bulk of PPh₃, the proposed structures of the two complexes are presented in Figure 4. Moreover, the spectra also present the ³¹P NMR signals at -5.18 ppm for complex **4** and -5.05 ppm for complex **6** which are attributed to the presence of unreacted PPh₃. Such the peak was not observed in complex **2** according to the purified single crystal collected for the analysis by ³¹P NMR spectroscopic technique. The ³¹P spectra of complexes **2**, **4** and **6** are presented in Figure 5.



Figure 4. The proposed structures of (a) $[Ni(ace)_2(PPh_3)_2]$ (4) and (b) $[Cu(ace)_2(PPh_3)_2]$ (6).



Figure 5. ³¹P spectra of $[Ag(ace)(PPh_3)]$ (2), $[Ni(ace)_2(PPh_3)_2]$ (4) and $[Cu(ace)_2(PPh_3)_2]$ (6).

The orientation of PPh₃ on the nickel(II) and copper(II) metal centers is supposed to be similar according to the comparable in their chemical shifts. Moreover, these values are also different from silver(I) complex in which the structure is in a dimer fashion with trigonal planar around the metal center (Figure 3). Unfortunately, for ¹H NMR study, the spectra could not be obtained to verify the complex formation as the signals of protons in acesulfame are all obscured by abundant protons of phenyl rings in PPh₃. For the complexes contained only acesulfame ligand; complexes **1**, **3** and **5**, ¹H NMR study could be constructed. The ¹H spectra of the novel complex **5** revealed the signals at the chemical shifts of 5.26 and 1.89 ppm belonged to the proton at *para* position of sulfone functional group (H_A) and the protons of methyl substituent (H_B) with the integration ratio of 1:3 (Figure 6). These peaks exhibited upfield shifted from 6.04 and 2.20 ppm in free acesulfame ion. Additionally, ¹H NMR of known complexes **1** and **3** also provided similar results to the literature (Icbudak et al., 2006; Cavicchioli et al., 2010).



Figure 6. Structure of acesulfame anion.

Mass spectrometry studies

Mass spectra for novel complexes $[Ni(ace)_2(PPh_3)_2](4)$, $[Cu(ace)_2(H_2O)_2](5)$ and $[Cu(ace)_2(PPh_3)_2](6)$ were examined the m/z⁺ ratio in order to verify the complex formation. $[Cu(ace)_2(PPh_3)_2](6)$ exhibits the peak at 911.8188 which exactly corresponding to the molecular mass of this substrate. Complex $[Ni(ace)_2(PPh_3)_2](4)$, the molecular mass of this complex is committed to be 906.09, unfortunately; the peak was presented at 857.1539. The observed signal might be attributed to the leaving of SO₂ group in which the result is comparable to the previous report by Yurdakul and Kose (2014). Moreover, complexes 4 and 6 also revealed the peaks at 587.2127 and 583.2746 belonged to the metal center bound to two molecules of PPh₃ as a minor peak. $[Cu(ace)_2(H_2O)_2](5)$ displays m/z⁺ signal at 525.0061 which is not as same as the suggested molecular structure of the desired product. Therefore, the molecular structure of complex 5 is proposed to be dinuclear complex with acesulfame as bridging ligands as the structure shown in Figure 7.



Figure 7. The proposed structure of complex $[Cu(ace)_2(H_2O)_2]$ (5).

X-Ray powder diffraction studies

The powder X-ray diffraction studies was carried with CuK_{α} (1.54 Å) radiation for $[Ni(ace)_2(PPh_3)_2](4)$, $[Cu(ace)_2(H_2O)_2](5)$ and $[Cu(ace)_2(PPh_3)_2](6)$ complexes. The X-ray diffraction pattern of the complexes representing in Figure 8, 9 and 10, respectively and d-spacing corelated to θ are given in Table 3. The average crystalline sizes of the complexes were calculated using Debye Scherrer equation (D = K λ/β cos θ) (D = Particle size, K = Dimensionless shape factor, λ = X-ray wavelength (0.154 Å) β = full width at half maximum of the diffraction peak, θ = Diffraction angle) (Manjuraj, 2018).



Figure 8. X-ray diffraction patterns of [Ni(ace)₂(PPh₃)₂](4).



Figure 9. X-ray diffraction patterns of $[Cu(ace)_2(H_2O)_2](5)$.



Figure 10. X-ray diffraction patterns of [Cu(ace)₂(PPh₃)₂](6)

Peak	[Ni(ace) ₂ (PPh ₃) ₂](4)		[Cu(ace) ₂ (H ₂ O) ₂](5)			[Cu(ace) ₂ (PPh ₃) ₂](6)			
no.	2 0	sin 0	d	2 0	sin 0	d	20	sin 0	d
1	5.69	0.0496	15.513	8.36	0.0729	10.562	8.66	0.0755	10.198
2	15.41	0.1341	5.743	12.70	0.1106	6.962	8.90	0.0776	9.923
3	18.12	0.1575	4.890	16.81	0.1462	5.267	9.39	0.0819	9.402
4	19.45	0.1689	4.558	19.03	0.1653	4.658	9.71	0.0847	9.091
5	23.57	0.2042	3.770	25.36	0.2195	3.508	19.37	0.1681	4.581
6	35.62	0.3059	2.517	25.42	0.2200	3.500	20.90	0.1814	4.245

Table 3. XRD data of complexes 4-6.

Anticancer studies

The prepared complexes were examined in the anticancer activity against A549 lung cancer cells and compared with starting materials and the standard Etoposide. The MTT assay of some examined substrates from this study is presented in Figure 11.



Figure 11. MTT assay of (a) NiCl₂(PPh₃)₃, Ni(CH₃CO₂)₂·6H₂O and Cu(CH₃CO₂)₂·xH₂O (b) complexes 4, 5 and 6.

The IC₅₀ values of complexes **1**, **3** and **5** which contained only accsulfame ligand are 42.04, 50.79 and 222.40, respectively. The results indicate deficient ability of such complexes in comparison to the standard compound, however; these values are still superior than their starting materials. Therefore, this experiment could be confirmed that the synthesized complexes with the introduction of acesulfame performed better activity than non-coordinated compounds and their metal precursors. The excellent results were achieved by complexes with the mixed ligands of acesulfame and triphenylphosphine compounds. Complexes **2**, **4** and **6** provided the IC₅₀ values of 1.65, 13.45 and 1.06, respectively. Amongst the studied metal centers, copper(II) of complex **6** exhibited the best inhibition against the growth of A549 cancer cells. The obtained results are proposed to be involved the influence of metal type and also the proportions of the mixed ligands on the prepared complexes. However, the effect of mixed ligands ratio and structural geometry on biological activity should be further investigated. The results of all biological studies are shown in Table 4.

Compounds	IC ₅₀ (μM)	Compounds	IC ₅₀ (μM)	
acesulfame	N/A*	PPh ₃	N/A	
AgNO ₃	N/A	Ni(CH ₃ CO ₂) ₂ ·4H ₂ O	103.70	
NiCl ₂ (PPh ₃) ₃	24.95	Cu(CH ₃ CO ₂) ₂ ·xH ₂ O	436.70	
$[Ag(ace)]_n$ (1)	42.04	$[Ag(ace)(PPh_3)]$ (2)	1.65	
$[Ni(ace)_2(H_2O)_4]$ (3)	50.79	$[Ni(ace)_2(PPh_3)_2]$ (4)	13.45	
$[Cu(ace)_2(H_2O)_2](5)$	222.40	$[Cu(ace)_2(PPh_3)_2]$ (6)	1.06	
Etoposide	5.62			

Table 4. IC_{50} values of the studied compounds.

Note: * The IC₅₀ value cannot be determined.

DISCUSSION

Analysis of the synthesized complexes 1-6 using spectroscopic methods including X-Ray powder diffraction could verify formation of the desired products. Complex $[Ag(ace)]_n$ (1) and $[Ni(ace)_2(H_2O)_4](3)$ have been already published in their molecular structures and the results from this study are comparable to the previous report. Complex $[Ag(ace)(PPh_3)](2)$ was synthesized established on the literature that using of mixed ligands would increase ability of complexes in the growth inhibition of cancer cells. The previous report by our group revealed the molecular structure obtained from single crystal

X-ray crystallography which demonstrated the dinuclear complex with trigonal planar structure around silver(I) ion. $[Ni(ace)_2(PPh_2)_2]$ (4), $[Cu(ace)_2(H_2O)_2]$ (5) and $[Cu(ace)_2(PPh_1)_2]$ (6) were originally prepared in this research. The results from spectroscopic techniques including X-Ray powder diffraction revealed the formation of theses complexes. $[Ni(ace)_2(PPh_3)_2]$ (4) exhibited the change in its ³¹P chemical shift up to 25.62 ppm compared to -5.25 ppm of the free PPh₂. Although, the mass spectrum is not corresponding to the proposed structure, the obtained result is still similar to the previous research in which the complex is supposed to lose its SO₂ molecule upon the investigation. Copper(II) complex in the presence of only acesulfame ligand (complex 5) revealed the mass spectrum at the peak corelated to the dinuclear structure as presented in Figure 7. IR study exhibits the coordination of water and acesulfame molecules towards the metal center. Additionally, the ¹H NMR spectrum also displays the change in chemical shifts of proton at *para* position of sulfone group and protons of methyl substituent compared to the free acesulfame which could confirm the complex formation. Both spectroscopic studies and mass spectrometry results confirm the formation of complex $[Cu(ace), (PPh_2)]$ (6) with the structure shown in Figure 4. The X-ray powder diffraction studies showed that $Cu(ace)_2(H_2O)_2(5)$ exhibited sharp peaks indicating high crystallinity of the complex. Based on literature, this may be according to the incorporation of water molecules into the coordination sphere (Khan, 2013). The average crystalline sizes of the complexes calculated using Debye Scherrer equation of [Ni(ace)₂(PPh₂)₂](4), [Cu(ace)₂ (H₂O)₂](5) and [Cu(ace)₂(PPh₂)₂](6) are 34.02, 31.50 and 23.86 nm, respectively suggesting the nanocrystalline phase of all the complexes. (Manjuraj, 2018)

The biological studies display the effect of mixed ligands towards the inhibition of A549 lung cancer cells growing. Starting materials provide high IC_{50} values which indicate unsatisfactory ability in the interaction with biological substrates and some cannot be even evaluated the IC_{50} values. The studies demonstrate the importance of coordination compounds in the biological study in which the use of copper(II) complex gave the most excellent capability for killing those cancer cells. The influence of mixed ligand ratio and also geometric structures should be further investigated.

CONCLUSION

In summary, metal complexes with account and in the absence and in the presence of triphenylphosphine were prepared and characterized. The biological study with cancer cells revealed great activity of the synthesized complexes in comparison to their starting substrates. The capability is increased when

triphenylphosphine was introduced to the metal complexes. Anyhow, the effect of the proportions of acesulfame and triphenylphosphine including the exact molecular structures of the synthesized complexes should be further investigated.

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